

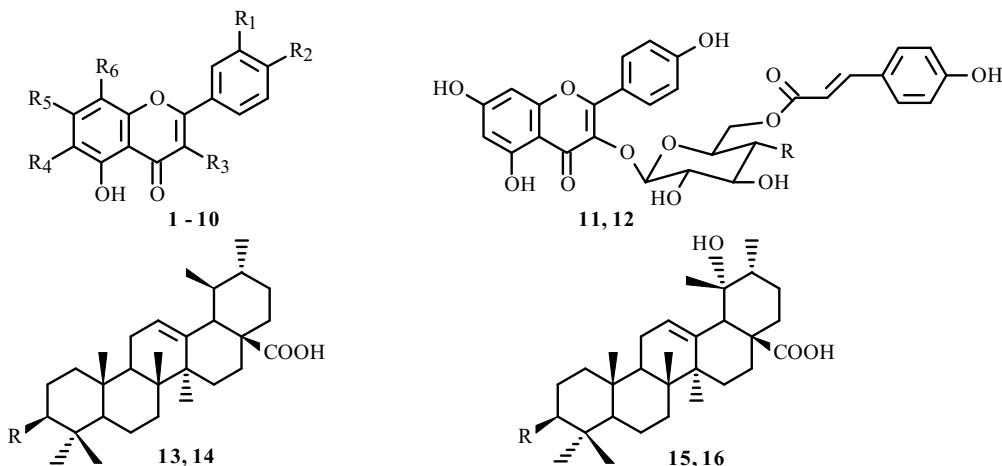
## FLAVONOIDS AND TRITERPENOIDS FROM *Anaphalis margaritacea*

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We have carried out systematical phytochemical studies on several medicinal herbs used in traditional Tibetan medicine growing only at 2500–4000 m above sea level on Qinghai-Tibet Plateau and found that all these plants are abundant in flavonoids [1–5]. The genus *Anaphalis* (Compositae) is widely distributed in the west and southwest regions of China, and 17 species of them are used as medicinal plants. *Anaphalis margaritacea* (Linn.) Benth. et Hook. f. has long been used by local inhabitants as a Tibetan medicine for cough and respiratory problems as well as for colds and rheumatism. Previous chemical investigation of this plant only revealed the presence of flavonoids, polyacetylenes, and hydroxylactone [6–9].

Herein we report the results of a thorough phytochemical study on flavonoids and triterpenoids from the whole plant of *Anaphalis margaritacea* (Linn.) Benth. et Hook. f. and their free-radical-scavenging potentials. Isolation of various fractions of the ethanol extract by column chromatography gave 12 known flavonoids. They were apigenin (**1**) [10], quercetin (**2**) [2, 8], kaempferol-3-O-β-D-glucopyranoside (**3**) [9], 3-methylquercetin (**4**) [5], spiraein (**5**) [10], 5,7-dihydroxy-3-methoxyflavone (**6**) [11], 5,7-dihydroxy-3,6,8-trimethoxyflavone (**7**), 5,7-dihydroxy-3,8-dimethoxyflavone (**8**), 5,6-dihydroxy-3,7-dimethoxyflavone (**9**) [4], 3,5-dihydroxy-6,7,8-trimethoxyflavone (**10**), tiliroside (**11**) [5] and kaempferol-3-O-[6''-O-(*trans*-*p*-coumaroyl)-4''-O-acetyl]-β-D-glucopyranoside (**12**) [12], together with four known triterpenoids, ursolic acid (**13**) [13], 3β-O-acetylursolic acid (**14**) [14], polnolic acid (**15**) [15], and 19α-hydroxy-3-acetylursolic acid (**16**) [16]. Their structures were established by direct comparison of their spectroscopic data with the corresponding literature values. Compounds **1–3**, **5**, **6**, **12–16** were isolated from *A. margaritacea* for the first time.



- 1:** R<sub>1</sub> = R<sub>3</sub> = R<sub>4</sub> = H, R<sub>2</sub> = R<sub>5</sub> = OH; **2:** R<sub>1</sub> = R<sub>2</sub> = R<sub>3</sub> = R<sub>5</sub> = OH, R<sub>4</sub> = R<sub>6</sub> = H  
**3:** R<sub>1</sub> = R<sub>4</sub> = R<sub>6</sub> = H, R<sub>2</sub> = R<sub>5</sub> = OH, R<sub>3</sub> = OGlc; **4:** R<sub>1</sub> = R<sub>2</sub> = R<sub>5</sub> = OH, R<sub>3</sub> = OMe, R<sub>4</sub> = R<sub>6</sub> = H  
**5:** R<sub>1</sub> = OGlc, R<sub>2</sub> = R<sub>3</sub> = R<sub>5</sub> = OH, R<sub>4</sub> = R<sub>6</sub> = H; **6:** R<sub>1</sub> = R<sub>2</sub> = R<sub>4</sub> = R<sub>6</sub> = H, R<sub>3</sub> = OMe, R<sub>5</sub> = OH  
**7:** R<sub>1</sub> = R<sub>2</sub> = H, R<sub>3</sub> = R<sub>4</sub> = R<sub>6</sub> = OMe, R<sub>5</sub> = OH; **8:** R<sub>1</sub> = R<sub>2</sub> = R<sub>4</sub> = H, R<sub>3</sub> = R<sub>6</sub> = OMe, R<sub>5</sub> = OH  
**9:** R<sub>1</sub> = R<sub>2</sub> = R<sub>6</sub> = H, R<sub>3</sub> = R<sub>4</sub> = OMe, R<sub>5</sub> = OH; **10:** R<sub>1</sub> = R<sub>2</sub> = H, R<sub>3</sub> = OH, R<sub>4</sub> = R<sub>5</sub> = R<sub>6</sub> = OMe  
**11, 13, 15:** R = OH, **12, 14, 16:** R = OAc

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TABLE 1. Free Radical Scavenging Activity of Isolated Compounds and VE in the DPPH Assay

Compound	IC <sub>20</sub> (μM)	Compound	IC <sub>20</sub> (μM)
<b>1</b>	>100	<b>7</b>	>100
<b>2</b>	6.36±0.11	<b>8</b>	>100
<b>3</b>	>100	<b>9</b>	>100
<b>4</b>	7.52±0.10	<b>10</b>	40.90±0.23
<b>5</b>	11.83±0.11	<b>11</b>	>100
<b>6</b>	>100	<b>12</b>	>100
VE	7.60		

Results are given as mean ±SD. Statistical analyses were performed using Student's *t*-test.

For the evaluation of free-radical-scavenging properties of compounds **1–12**, a DPPH assay was adopted with vitamin E (VE) as positive control. In the DPPH system, the free radical scavenging activity of tested samples was expressed as IC<sub>20</sub>. The results of scavenging activities of compounds **1–12** and VE are listed in Table 1; **2** (IC<sub>20</sub>= 6.36±0.11 μM) and **4** (IC<sub>20</sub>= 7.52±0.10 μM) was most active in the tested compounds. The higher scavenging activities were associated with the *ortho*-dihydroxyl B-ring structures in the flavonoids, conferring higher radical stability and participating in electron delocalization, as reported for other systems [2, 17].

**General.** The <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on Varian Mercury-300BB NMR (300 MHz) spectrometers with TMS as internal standard. Silica gel (200–300 mesh) used for column chromatography (CC) and silica gel GF<sub>254</sub> (10–40 μm) used for TLC were supplied by Qingdao Marine Chemical Factory, Qingdao, P. R. China. VE was purchased from Aldrich, USA and DPPH from Sigma, USA.

**Plant Material.** The whole plant of *Anaphalis margaritacea* (Linn.) Benth. et Hook. f. was collected in Huzhu County, Qinghai province, P. R. China, in August 2005. It was identified by Dr. Huan-Yang Qi. The voucher specimens (2005A002) were deposited in the Key Laboratory for Natural Medicine of Gansu Province, Lanzhou Institute of Chemical Physics, Chinese Academy of Sciences, P. R. China.

**Extraction and Isolation.** Powdered and dried whole plant of *A. margaritacea* (2.6 kg) was extracted with 95% EtOH at room temperature. The combined extract (183 g) was then suspended in H<sub>2</sub>O (1.0 L) and extracted successively with petroleum ether, EtOAc, *n*-BuOH (3 × 1.0 L each), respectively. The solvents were distilled to produce the petroleum ether fraction (40 g), the EtOAc fraction (43 g), and the *n*-BuOH fraction (26 g).

The petroleum ether extract was chromatographed on a silica gel (200–300 mesh, 450 g, 8 × 140 cm) column using petroleum ether–EtOAc. Elution gave six compounds, namely **6** (17.6 mg), **8** (21.3 mg), **9** (14.8 mg), **10** (12.4 mg), **14** (13.7 mg), and **16** (15.2 mg). The ethyl acetate extract was chromatographed on a silica gel (200–300 mesh, 500 g, 8 × 140 cm) column using petroleum ether–acetone. Elution of the column gave 10 compounds, namely **1** (21.3 mg), **2** (17.1 mg), **3** (12.4 mg), **4** (16.8 mg), **5** (24.8 mg), **7** (18.7 mg), **11** (30.3 mg), **12** (31.5 mg), **13** (19.6 mg), and **15** (12.7 mg).

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